



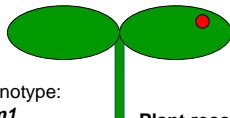
The effect of rotation of R gene on frequencies of avirulence alleles in blackleg populations

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### Major gene resistance is a gene-for gene interaction

- Plant contains resistance genes (R genes)
- Pathogen contains avirulence genes (Avr genes) corresponding to R genes



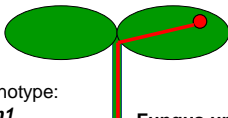
Plant Genotype: *Rlm1*

Fungus Genotype: *AvrRml1* (Avirulent)

Plant recognises fungus  
- Defence mechanisms  
- **NO INFECTION**

### Major gene resistance is a gene-for gene interaction





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Plant Genotype: *Rlm1*

Fungus Genotype: *avrRml1* (Virulent)

Fungus undetected by plant  
**INFECTION/DISEASE**

|  | cv. AV-Garnet<br>( <i>Rlm1</i> )  | cv. CB Telfer<br>( <i>Rlm4</i> )   |
|--|---|--|
| D13<br>( <i>avrLm1</i> ; <i>AvrLm4</i> ) |   |   |
| D14<br>( <i>AvrLm1</i> ; <i>avrLm4</i> ) |  |  |

### Take home messages

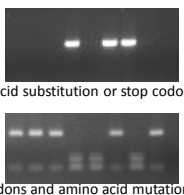
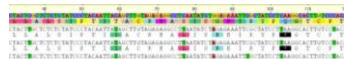
- Monitoring changes in frequencies of avirulence alleles using molecular markers
  - Enables risk of resistance breakdown to be determined
- Rotation of cultivars with different R genes can influence allele frequencies
  - Affects both the corresponding avirulence gene and linked avirulence genes
- Rotations of some R genes will have more benefit than others
- Genomic location (linkage) of R genes in plant and Avr genes in pathogens may influence selection pressure on pathogen by sowing particular R genes

### Five avirulence genes have been cloned from *L. maculans*

- AvrLm1* - *Rlm1* (AV-Garnet, Group A)
- AvrLm4-7* - *Rlm4* (CB Telfer, Group B)
  - *Rlm7* (not in current cultivars)
- AvrLm2* - *Rlm2* (presence unknown in current cultivars)
- AvrLm5* - *Rlm5* (Juncea R gene)
- AvrLm6* - *Rlm6* (Juncea R gene)

### Deletion, point mutations and RIP mutation confer virulence towards R genes

- Different mutation events occur in different avirulence genes.
  - Can develop molecular markers to screen isolates to determine virulence genotype.
- Deletion of entire gene
  - Presence/absence PCR screen
    - AvrLm1*
    - AvrLm6*
- Single point mutation (confers amino acid substitution or stop codon)
  - PCR and restriction digest
    - AvrLm4*
    - AvrLm2*
    - AvrLm5*
- RIP mutations (generate many stop codons and amino acid mutations)
  - Sequence gene
    - AvrLm7*

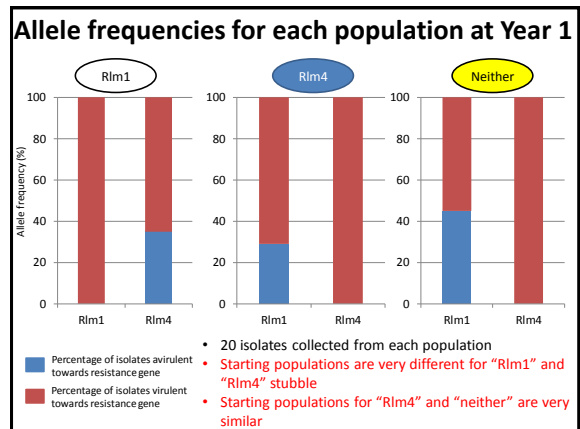
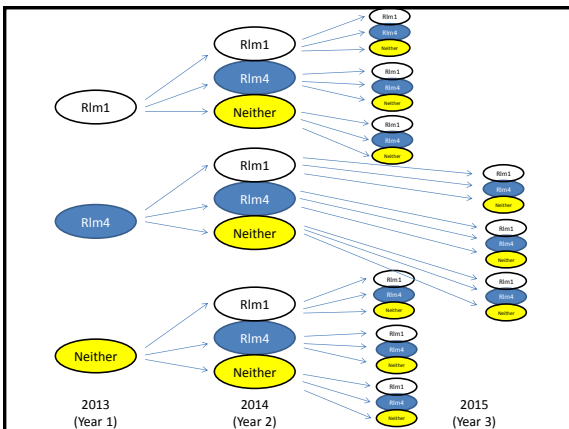
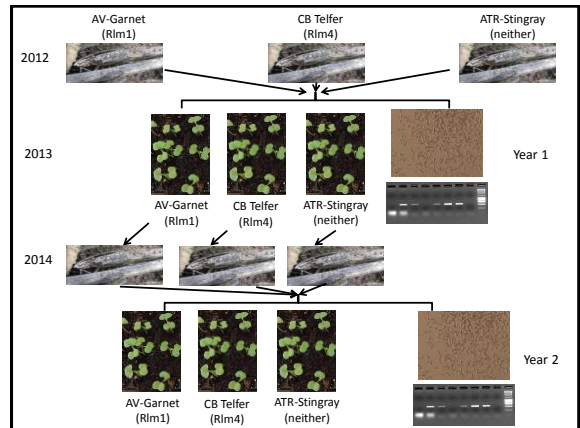



### Molecular markers used to monitor changes in avirulence allele frequencies

- Blackleg isolates (ascospores) collected from stubble since 2000
- DNA prepared and tested with molecular markers for each avirulence gene
  - 1901 isolates in total
    - 657 from Vic
    - 427 from NSW
    - 675 from SA
    - 142 from WA
- These data used to confirm disease severity in field and glasshouse experiments
  - Supported the findings that R gene rotation influences Avr allele frequencies.
  - However, isolates always selected randomly from different locations under different environmental conditions

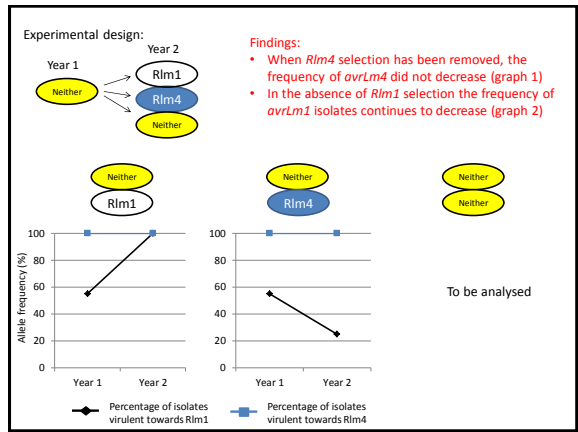
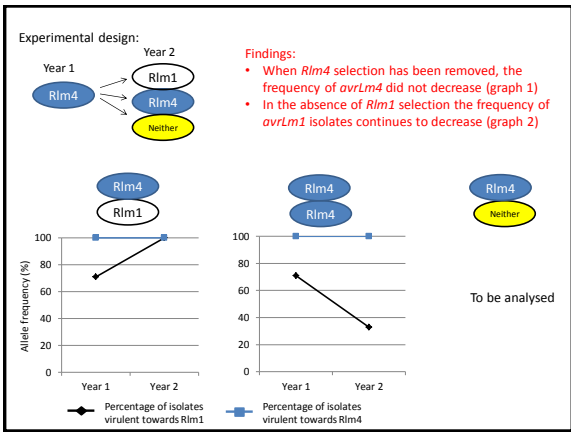
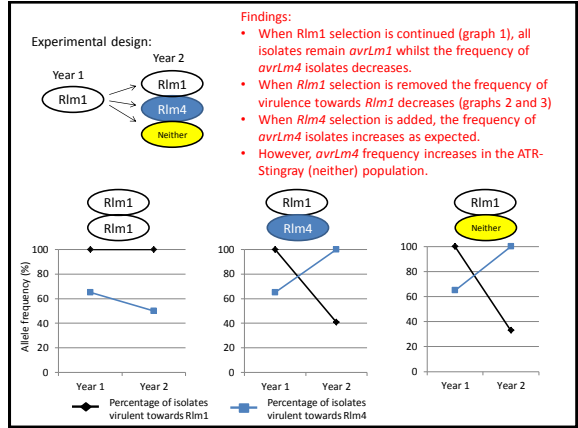
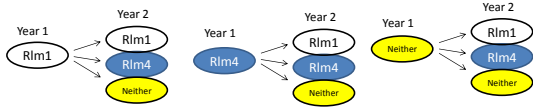
### 3 year experiment to monitor effect of cultivar rotation on allele frequencies

- Recurring selection experiment established in 2012
- Cultivars with known R genes (*Rlm1*, *Rlm4* or neither)
- Stubble releases ascospore inoculum onto seedlings in shadehouse experiments
- Frequency of alleles of *AvrLm1* and *AvrLm4* determined by high through put molecular assay



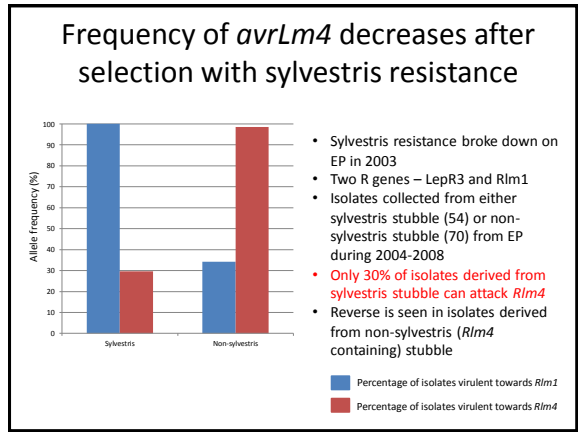
### Allele frequencies for each population at year 2

- 20 isolates collected from each selection regime (year 2 population)
- Analysed for virulence towards *Rlm1* and *Rlm4*

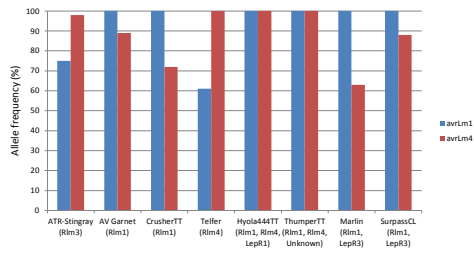


### Changes in allele frequencies differ for different avirulence genes

- Changes to *avrLm1* allele frequencies as expected (decreased when *Rlm1* selection removed)
- Frequency of *avrLm4* isolates did not decrease when *Rlm4* selection was removed
  - No fitness cost to the pathogen?
  - Changes in frequency of this avr gene are slow to occur?
  - Know that *avrLm4* frequency can change with rotation (sylvestris situation)

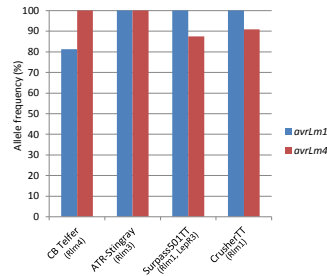


## Similar differences in allele frequency are seen in a single year



577 isolates collected from stubble of different cultivars across Australia in 2014

## Allele frequencies collected at a single site can vary



- 10 isolates cultured from stubble of various cultivars collected from Parkes, NSW
- Allele frequencies for both *AvrLm1* and *AvrLm4* vary, depending on the stubble from which isolates are cultured
- Findings similar to experiments using ascospore shower technique

## Changes in allele frequencies may depend on gene location in the host or pathogen

- Blackleg populations derived from stubble of ATR-Stingray (neither *Rlm1* or *Rlm4*) behaved very similarly to those derived from *Rlm4* stubble
  - ATR-Stingray has *Rlm3*
  - *Rlm3-4-7-9* are linked in the plant
  - *AvrLm3-4-7-9* are linked in the fungus
- Does gene location influence Avr allele frequencies?
  - *AvrLm1* and *AvrLm6* are genetically linked in the pathogen
  - *LepR3* (*Rlm1*?), *Rlm2* and *Rlm6* are reported as alleles of the same gene (Larkan et al.)
  - Selection from sylvestris cultivars (*Rlm1*) influences frequency of *AvrLm6* in blackleg populations (Van de Wouw et al. 2010).

## Conclusions

- Rotation of cultivars with different resistance genes influence frequencies of avirulence genes
- Not all avirulence genes respond similarly to this rotation
- Need to understand the interaction between Avr genes and R genes better
  - Linkage of Avr or R genes influences allele frequencies
  - Some rotations will have more benefit than others e.g. *LepR3* followed by *Rlm4*
- We still don't know
  - What frequency of virulent isolates leads to resistance being overcome?
  - Whether we should rotate R genes every year or every 3 years?