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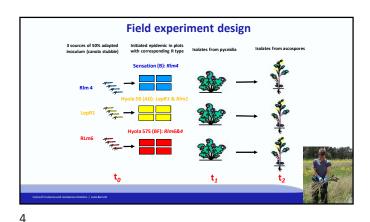
Motivation for empirical and modelling experiments

1. What mechanisms underlie the ability of rotations to control disease?

2. How do pathogen populations respond to different resistance combinations at different phases in the epidemic cycle?

3. What is the optimal strategy to rotate major gene resistance?

Within host competition



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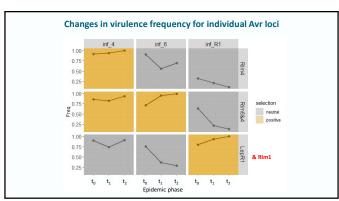
Pathogen population composition

Pathogen isolates obtained from inoculum and leaf-spot populations (~2500 isolates)
Phenotyping for infectivity of ~1500 isolates on RIm4, RIm6 and LepR1

Construct multi-locus infectivity phenotypes

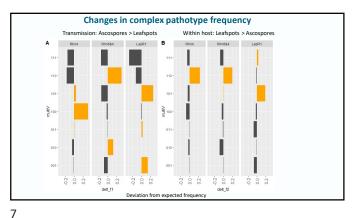
RIM4 # RIm6 # LepR1
0/1 0/1 0/1
000, 001, 010, 011, 100, 101, 110, 111

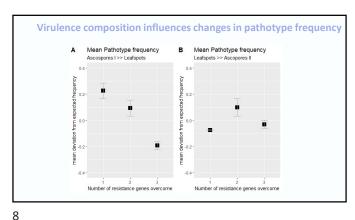
Estimates of virulence and pathotype frequency at t₀, t₁ and t₂



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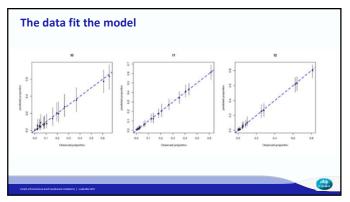


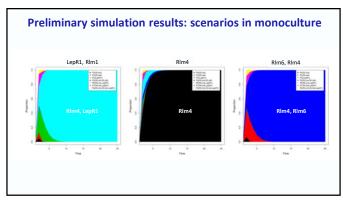


Canola-Blackleg: life-cycle genetic dynamics Strong evidence for selection against unnecessary virulence during transmission Selection for/against virulence changes during withinhost phase Indicates that costs of virulence vary at different epidemic phases What is the optimal strategy to rotate different R-types?

Modelling rotations 3 cultivars, 8 pathogen genotypes $\beta\text{:}$ Pathotype transmission rate. Estimate β based on change in frequency from to t1 Plants of cultivar *i* with leafspot infections from pathotype *k* γ: Pathotype reproductive rate. Estimate based on change in frequency from t₁ to t₂ Plants of cultivar *i* producing ascopsores of pathotype K

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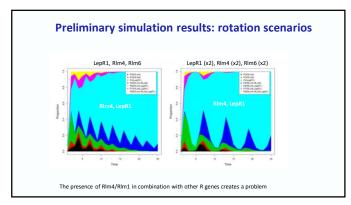




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Conclusions

- More modelling work is required
- More data required (other R genes, QR etc)
- Rotations offer potential for long-term disease control even if resistance has broken down.
- Optimal strategies will depend on many factors, e.g.:
- Costs of virulence
- Number of R-genes/cultivars
- Spatial factors
- Overcome resistance genes should not be deployed in combination with other R genes

Costs of virulence and resistance rotations | Luke Barrett

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Thanks to the following individuals for material, discussion and ideas:

Pete Thrall, Jeremy Burdon, Loup Rimbaud, Mathieu Legros, Susie Sprague, Kristy Lam — CSIRO
Lydia Bousset, Julien Papalx—INRA, France
Funding: CSIRO, INRA and the GRDC

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